

PLNmodels

A collection of Poisson lognormal models
for multivariate analysis of count data

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<https://pln-team.github.io/PLNmodels>

Reproducibility

R/C++ Package PLNmodels

Last stable release on CRAN, development version available on GitHub.

```
install.packages("PLNmodels")
devtools::install_github("PLN-team/PLNmodels")

library(PLNmodels)
packageVersion("PLNmodels")

## [1] '0.11.4'
```

Python module

A Python + PyTorch implementation is coming

Advertisement (more, sorry about that)

<https://computo.sfds.asso.fr>, a new journal promoting reproducible research

Resources

Help and documentation

The [PLNmodels website](#) contains the standard package documentation and a set of comprehensive vignettes for the top-level functions

Publications

Chiquet, J., M. Mariadassou, and S. Robin (2018). "Variational inference for probabilistic Poisson PCA". In: *The Annals of Applied Statistics* 12, pp. 2674-2698. URL: <http://dx.doi.org/10.1214/18-AOAS1177>.

Chiquet, J., M. Mariadassou, and S. Robin (2019). "Variational inference for sparse network reconstruction from count data". In: *Proceedings of the 19th International Conference on Machine Learning (ICML 2019)*.

Chiquet, J., M. Mariadassou, and S. Robin (2021). "The Poisson-Lognormal Model as a Versatile Framework for the Joint Analysis of Species Abundances". In: *Frontiers in Ecology and Evolution* 9. DOI: [10.3389/fevo.2021.588292](https://doi.org/10.3389/fevo.2021.588292).

Facon, B., A. Hafsi, M. C. de la Masselière, et al. (2021). "Joint species distributions reveal the combined effects of host plants, abiotic factors and species competition as drivers of species abundances in fruit flies". In: *Ecological Letters*. DOI: [10.1111/ele.13825](https://doi.org/10.1111/ele.13825).

Lejal, E., J. Chiquet, J. Aubert, et al. (2021). "Temporal patterns in Ixodes ricinus microbial communities: an insight into tick-borne microbe interactions". In: *Microbiome* 9:153. DOI: [10.1186/s40168-021-01051-8](https://doi.org/10.1186/s40168-021-01051-8).

Generic form of data sets

Routinely gathered in ecology/microbiology/genomics

Data tables

- Abundances: read counts of species/transcripts j in sample i
- Covariates: value of environmental variable k in sample i
- Offsets: sampling effort for species/transcripts j in sample i

Need a framework to model *dependencies between counts*

- understand environmental effects
 - ~~ explanatory models (multivariate regression, classification)
- exhibit patterns of diversity
 - ~~ summarize the information (clustering, dimension reduction)
- understand between-species interactions
 - ~~ 'network' inference (variable/covariance selection)
- correct for technical and confounding effects
 - ~~ account for covariables and sampling effort

Models for multivariate count data

If we were in a Gaussian world...

The general linear model [MKB79] would be appropriate! For each sample $i = 1, \dots, n$,

$$\underbrace{\mathbf{Y}_i}_{\text{abundances}} = \underbrace{\mathbf{x}_i^\top \boldsymbol{\Theta}}_{\text{covariates}} + \underbrace{\mathbf{o}_i}_{\text{sampling effort}} + \boldsymbol{\varepsilon}_i, \quad \boldsymbol{\varepsilon}_i \sim \mathcal{N}(\mathbf{0}_p, \underbrace{\boldsymbol{\Sigma}}_{\text{between-species dependencies}})$$

null covariance \Leftrightarrow independence \rightsquigarrow uncorrelated species/transcripts do not interact

\rightsquigarrow This model gives birth to Principal Component Analysis, Discriminant Analysis, Gaussian Graphical Models, Gaussian Mixture models and many others ...

With count data...

There is no generic model for multivariate counts

- Data transformation ($\log, \sqrt{}$): quick and dirty
- Non-Gaussian multivariate distributions [Ino+17]: do not scale to data dimension yet
- Latent variable models: interaction occur in a latent (unobserved) layer

The Poisson Lognormal model (PLN)

The PLN model [AH89] is a multivariate generalized linear model, where

- the counts \mathbf{Y}_i are the response variables
- the main effect is due to a linear combination of the covariates \mathbf{x}_i
- a vector of offsets \mathbf{o}_i can be specified for each sample.

$$\mathbf{Y}_i | \mathbf{Z}_i \sim \mathcal{P}(\exp \mathbf{Z}_i), \quad \mathbf{Z}_i \sim \mathcal{N}(\mathbf{o}_i + \mathbf{x}_i^\top \boldsymbol{\Theta}, \boldsymbol{\Sigma}),$$

The unknown parameters are

- $\boldsymbol{\Theta}$, the regression parameters
- $\boldsymbol{\Sigma}$, the variance-covariance matrix

Stacking all individuals together,

- \mathbf{Y} is the $n \times p$ matrix of counts
- \mathbf{X} is the $n \times d$ matrix of design
- \mathbf{O} is the $n \times p$ matrix of offsets

Properties: over-dispersion, arbitrary-signed covariances

- mean: $\mathbb{E}(Y_{ij}) = \exp(o_{ij} + \mathbf{x}_i^\top \boldsymbol{\Theta}_{\cdot j} + \sigma_{jj}/2) > 0$
- variance: $\mathbb{V}(Y_{ij}) = \mathbb{E}(Y_{ij}) + \mathbb{E}(Y_{ij})^2 (e^{\sigma_{jj}} - 1) > \mathbb{E}(Y_{ij})$
- covariance: $\text{Cov}(Y_{ij}, Y_{ik}) = \mathbb{E}(Y_{ij})\mathbb{E}(Y_{ik}) (e^{\sigma_{jk}} - 1).$

Natural extensions

Various tasks of multivariate analysis

- Dimension Reduction: rank constraint matrix Σ .

$$\mathbf{Z}_i \sim \mathcal{N}(\boldsymbol{\mu}, \Sigma = \mathbf{B}\mathbf{B}^\top), \quad \mathbf{B} \in \mathcal{M}_{pk} \text{ with orthogonal columns.}$$

- Classification: maximize separation between groups with means

$$\mathbf{Z}_i \sim \mathcal{N}(\boldsymbol{\mu}_k \mathbf{1}_{\{i \in k\}}, \Sigma), \quad \text{for known memberships.}$$

- Clustering: mixture model in the latent space

$$\mathbf{Z}_i \mid i \in k \sim \mathcal{N}(\boldsymbol{\mu}_k, \Sigma_k), \quad \text{for unknown memberships.}$$

- Network inference: sparsity constraint on inverse covariance.

$$\mathbf{Z}_i \sim \mathcal{N}(\boldsymbol{\mu}, \Sigma = \boldsymbol{\Omega}^{-1}), \quad \|\boldsymbol{\Omega}\|_1 < c.$$

- Variable selection: sparsity constraint on regression coefficients

$$\mathbf{Z}_i \sim \mathcal{N}(\mathbf{x}_i^\top \boldsymbol{\Theta}, \Sigma), \quad \|\boldsymbol{\Theta}\|_1 < c.$$

Inference: latent model but intractable EM

Estimate $\theta = (\Theta, \Sigma)$, predict the \mathbf{Z}_i , while the model marginal likelihood is

$$p_\theta(\mathbf{Y}_i) = \int_{\mathbb{R}^p} \prod_{j=1}^p p_\theta(Y_{ij}|Z_{ij}) p_\theta(\mathbf{Z}_i) d\mathbf{Z}_i$$

Maximum likelihood for incomplete data model: EM

With $\mathcal{H}(p) = -\mathbb{E}_p(\log(p))$ the entropy of p ,

$$\log p_\theta(\mathbf{Y}) = \mathbb{E}_{p_\theta(\mathbf{Z} | \mathbf{Y})} [\log p_\theta(\mathbf{Y}, \mathbf{Z})] + \mathcal{H}[p_\theta(\mathbf{Z} | \mathbf{Y})]$$

EM requires to evaluate (some moments of) $p_\theta(\mathbf{Z} | \mathbf{Y})$, but there is no close form!

Solutions

- [AH89] resort on numerical integration; [Kar05] Monte-Carlo integration
- Several heuristics, not always well motivated, found in the literature...
- Variational approach [WJ08]: use a proxy of $p_\theta(\mathbf{Z} | \mathbf{Y})$.

Variational approximation

Principle

- Find a proxy of the conditional distribution $p(\mathbf{Z} \mid \mathbf{Y})$:

$$q(\mathbf{Z}) \approx p_\theta(\mathbf{Z} \mid \mathbf{Y}).$$

- Choose a convenient class of distribution \mathcal{Q} and minimize a divergence

$$q(\mathbf{Z})^* \arg \min_{q \in \mathcal{Q}} D(q(\mathbf{Z}), p(\mathbf{Z} \mid \mathbf{Y})).$$

Popular choice

The Kullback-Leibler divergence (error averaged wrt the approximated distribution)

$$KL(q(\mathbf{Z}), p(\mathbf{Z} \mid \mathbf{Y})) = \mathbb{E}_q \left[\log \frac{q(z)}{p(z)} \right] = \int_Z q(z) \log \frac{q(z)}{p(z)} dz.$$

Variational EM & PLN

Class of distribution: diagonal multivariate Gaussian

$$\mathcal{Q} = \left\{ q : q(\mathbf{Z}) = \prod_i q_i(\mathbf{Z}_i), \quad q_i(\mathbf{Z}_i) = \mathcal{N}(\mathbf{Z}_i; \mathbf{m}_i, \text{diag}(\mathbf{s}_i \circ \mathbf{s}_i)) , \mathbf{m}_i, \mathbf{s}_i \in \mathbb{R}_p \right\}$$

Maximize the ELBO (Evidence Lower BOund):

$$J(\theta, q) = \log p_\theta(\mathbf{Y}) - KL[q_\theta(\mathbf{Z}) || p_\theta(\mathbf{Z} | \mathbf{Y})] = \mathbb{E}_q[\log p_\theta(\mathbf{Y}, \mathbf{Z})] + \mathcal{H}[q(\mathbf{Z})]$$

Variational EM

- VE step: find the optimal q (here, $\{(\mathbf{m}_i, \mathbf{s}_i)\}_{i=1,\dots,n} = \{\mathbf{M}, \mathbf{S}\}$):

$$q^h = \arg \max J(\theta^h, q) = \arg \min_{q \in \mathcal{Q}} KL[q(\mathbf{Z}) || p_{\theta^h}(\mathbf{Z} | \mathbf{Y})]$$

- M step: update $\hat{\theta}^h$

$$\theta^h = \arg \max J(\theta, q^h) = \arg \max_{\theta} \mathbb{E}_q[\log p_\theta(\mathbf{Y}, \mathbf{Z})]$$

ELBO and gradients for PLN

Let $\mathbf{A} = \mathbb{E}_q[\exp(\mathbf{Z})] = \exp\left(\mathbf{O} + \mathbf{M} + \frac{1}{2}\mathbf{S}^2\right)$

Variational bound

$$\begin{aligned} J(\mathbf{Y}) &= \mathbf{1}_n^\top ([\mathbf{Y} \circ (\mathbf{O} + \mathbf{M}) - \mathbf{A} + \log(\mathbf{S})]) \mathbf{1}_p + \frac{n}{2} \log |\boldsymbol{\Omega}| \\ &\quad - \frac{1}{2} \text{trace} \left(\boldsymbol{\Omega} \left[(\mathbf{M} - \mathbf{X}\boldsymbol{\Theta})^\top (\mathbf{M} - \mathbf{X}\boldsymbol{\Theta}) + \text{diag}(\mathbf{1}_n^\top \mathbf{S}^2) \right] \right) + \text{cst.} \end{aligned}$$

M-step (Analytical)

$$\hat{\boldsymbol{\Theta}} = (\mathbf{X}^\top \mathbf{X})^{-1} \mathbf{X} \mathbf{M}, \quad \hat{\boldsymbol{\Sigma}} = \frac{1}{n} (\mathbf{M} - \mathbf{X}\hat{\boldsymbol{\Theta}})^\top (\mathbf{M} - \mathbf{X}\hat{\boldsymbol{\Theta}}) + \frac{1}{n} \text{diag}(\mathbf{1}^\top \mathbf{S}^2)$$

Variational E-step (optimization)

$$\frac{\partial J(q)}{\partial \mathbf{M}} = (\mathbf{Y} - \mathbf{A} - (\mathbf{M} - \mathbf{X}\boldsymbol{\Theta})\boldsymbol{\Omega}), \quad \frac{\partial J(q)}{\partial \mathbf{S}} = \frac{1}{\mathbf{S}} - \mathbf{S} \circ \mathbf{A} - \mathbf{S} \mathbf{D}_{\boldsymbol{\Omega}}.$$

Application to the optimization of PLN models

Property of PLN variational approximation

The ELBO $J(\theta, q)$ is bi-concave, i.e.

- concave wrt $q = (\mathbf{M}, \mathbf{S})$ for given θ
- convex wrt $\theta = (\Sigma, \Theta)$ for given q but not jointly concave in general.

Optimization

Gradient ascent for the set of variational parameters (\mathbf{M}, \mathbf{S})

Medium scale problems

- **algorithm:** conservative convex separable approximations Svanberg [Sva02]
- **implementation:** `NLOpt` nonlinear-optimization library Johnson [Joh11]
- **initialization:** LM after log-transformation applied independently on each variables + concatenation of the regression coefficients + Pearson residuals

~~~ Comfortable up to a thousand of sites ( $n \approx 1000$ ), hundreds of species ( $p \approx 100s$ )

# Large scale problems

## Sophisticated Adaptive Stochastic Gradient Descent

- Rprop (1993) uses the gradient sign and update each variable independently:
- AdaGrad (2011) uses adaptive coordinate-wise step-sizes
- RMSProp (2012) adds momentum to the step-sizes
- Adam (2015) also adds momentum to the gradients



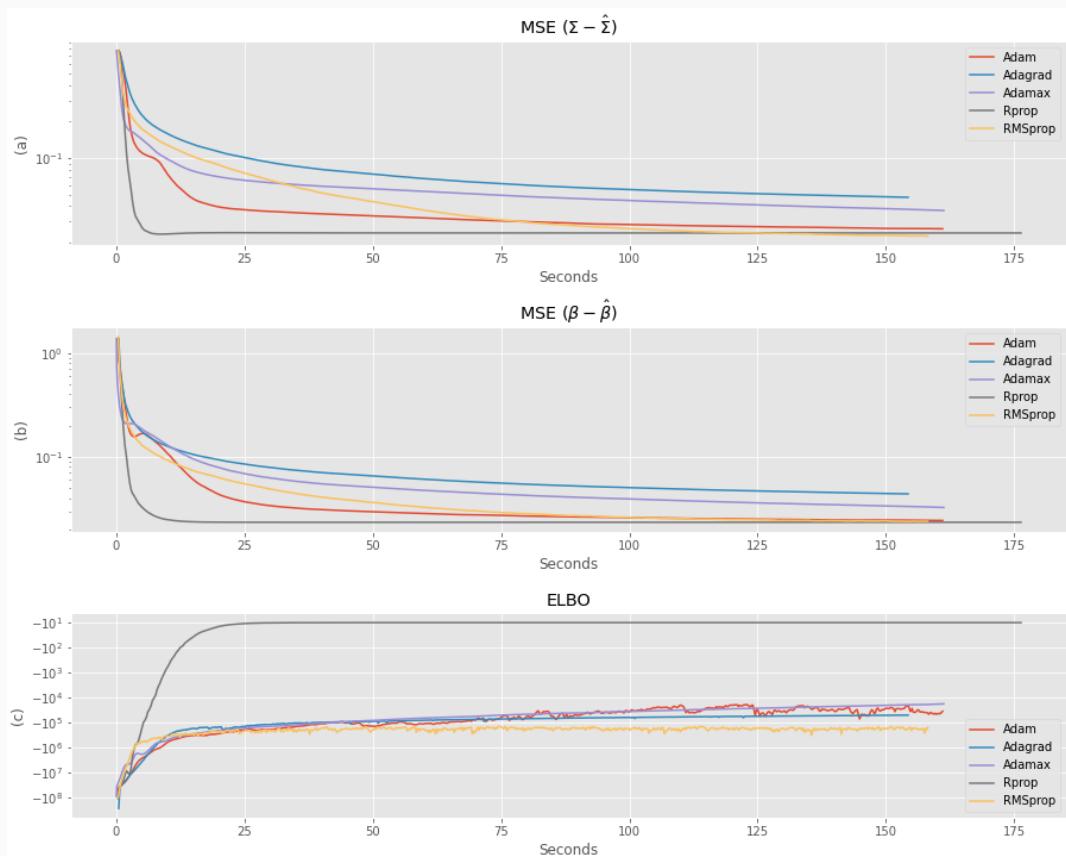
(a) SGD without momentum



(b) SGD with momentum

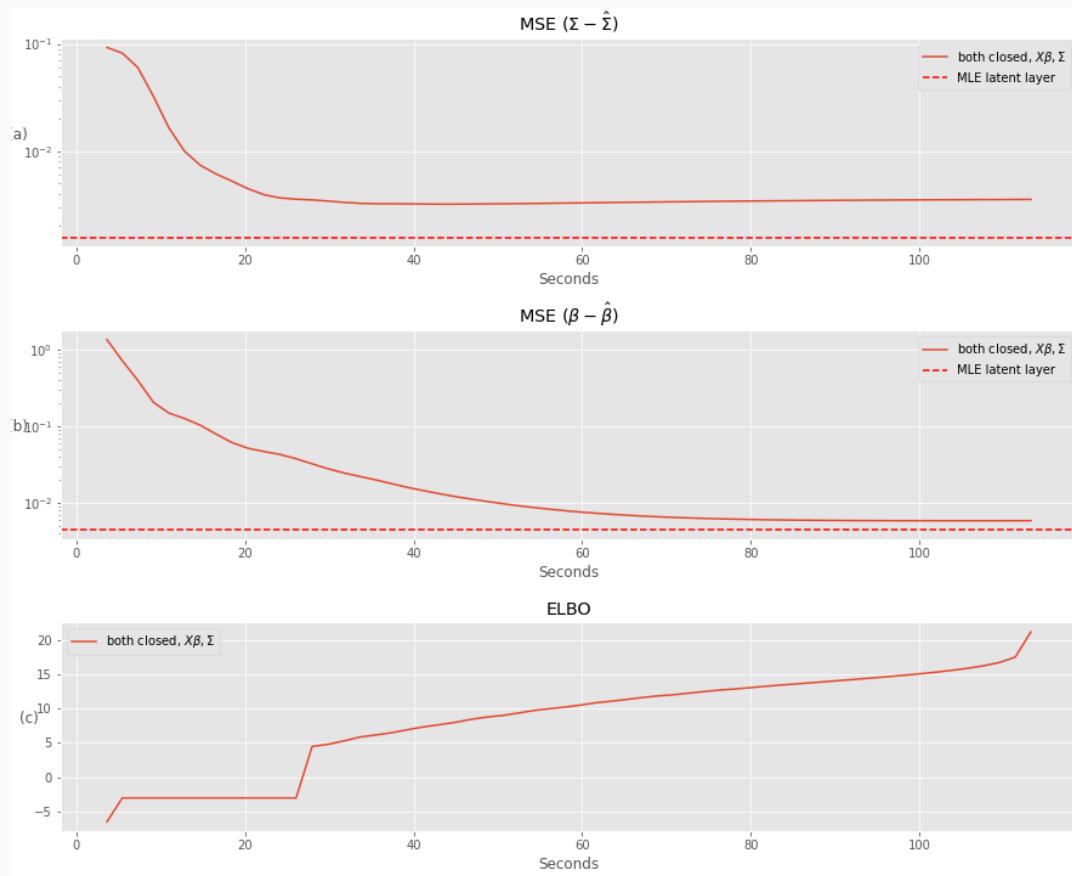
~~ All available in **Pytorch** with auto-differentiation.

# Optimizers comparison



$n = 1,000$ ,  $p = 200$ ,  $d = 2$ . Rprop is much faster.

# Performance



$n = 10,000, p = 2,000, d = 2$  (running time: 1 min 40s)

~~ Work up to  $n = 100,000, p = 10,000$ s (useful for microbiological data, single cell data, now found in ecology)

# Statistical Inference

## A first try: Wald test

Test  $\mathcal{H}_0 : R\theta = r_0$  with the statistic

$$(R\hat{\theta} - r_0)^\top \left[ nR\hat{\mathbb{V}}(\hat{\theta})R^\top \right]^{-1} (R\hat{\theta} - r_0) \sim \chi_k^2 \quad \text{where } k = \text{rank}(R).$$

The Fisher Information matrix

$$I(\hat{\theta}) = -\mathbb{E}_{\theta} \left[ \frac{\partial^2 \log \ell(\theta; x)}{\partial \theta^2} \right]$$

can be used as an approximation of  $n\hat{\mathbb{V}}(\hat{\theta})^{-1}$ .

## Application

Derive confidences interval for the inverse covariance  $\boldsymbol{\Omega}$  and the regression parameters  $\boldsymbol{\Theta}$ .

# Variational Wald-test

## Variational Fisher Information

The Fisher information matrix is given by

$$I(\theta) = \begin{pmatrix} \frac{1}{n} (\mathbf{I}_p \otimes \mathbf{X}^\top) \text{diag}(\text{vec}(\mathbf{A})) (\mathbf{I}_p \otimes \mathbf{X}) & \mathbf{0} \\ \mathbf{0} & \frac{1}{2} \boldsymbol{\Omega}^{-1} \otimes \boldsymbol{\Omega}^{-1} \end{pmatrix}$$

and can be inverted blockwise to estimate  $\hat{\mathbb{V}}(\hat{\theta})$ .

## Wald test and coverage

- $\hat{\mathbb{V}}(\boldsymbol{\Theta}_{kj}) = [n(\mathbf{X}^\top \text{diag}(\text{vec}(\hat{A}_{.j})) \mathbf{X})^{-1}]_{kk}$
- $\hat{\mathbb{V}}(\boldsymbol{\Omega}_{kl}) = 2\boldsymbol{\Omega}_{kk}\boldsymbol{\Omega}_{ll}$

The confidence intervals at level  $\alpha$  are given by

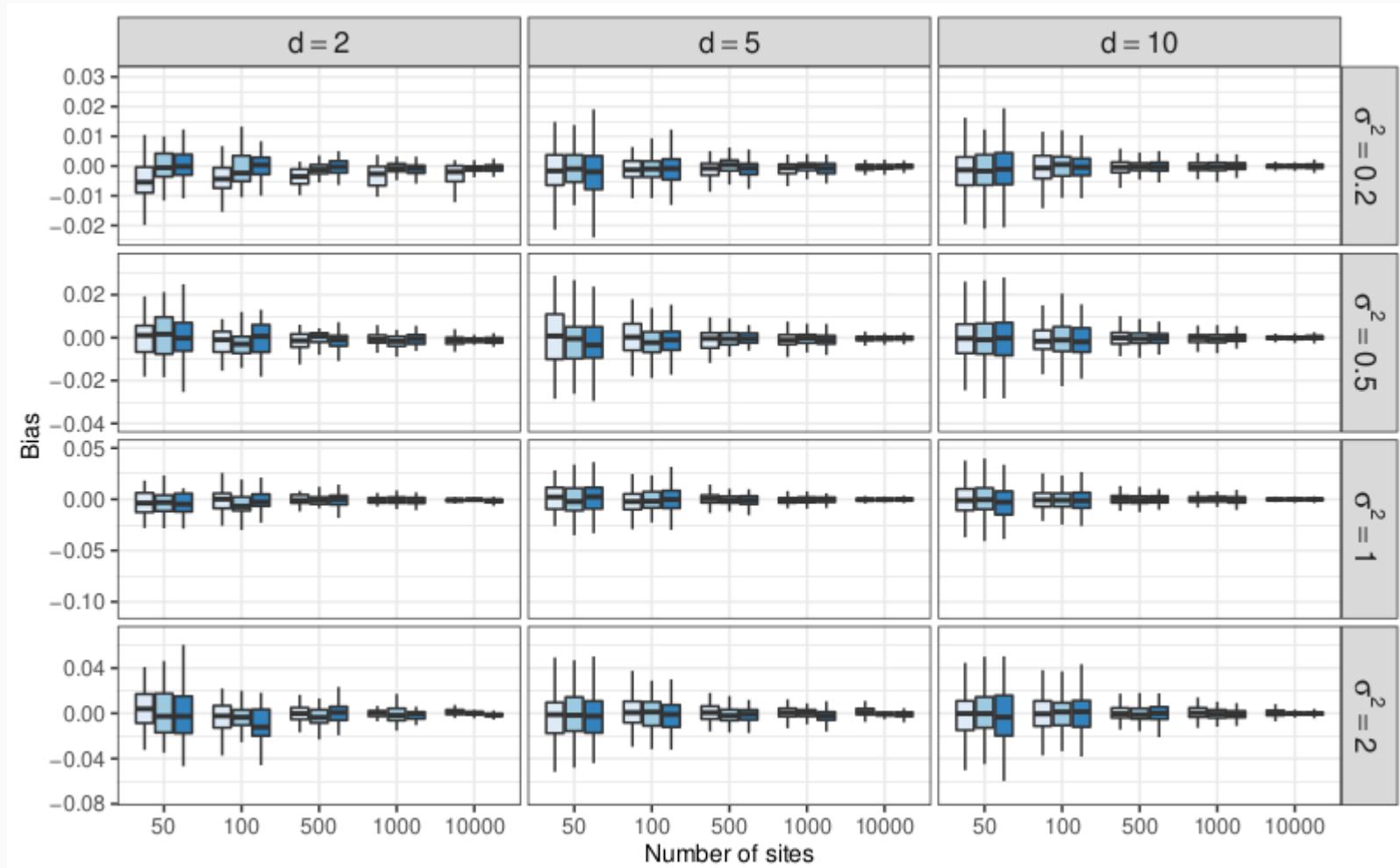
- $B_{kj} = \hat{B}_{kj} \pm \frac{q_{1-\alpha/2}}{\sqrt{n}} \sqrt{\hat{\mathbb{V}}(\boldsymbol{\Theta}_{kj})}$
- $\boldsymbol{\Omega}_{kl} = \hat{\boldsymbol{\Omega}}_{kl} \pm \frac{q_{1-\alpha/2}}{\sqrt{n}} \sqrt{\hat{\mathbb{V}}(\boldsymbol{\Omega}_{kl})}$ .

# Numerical study

## Study Bias and coverage of the estimator

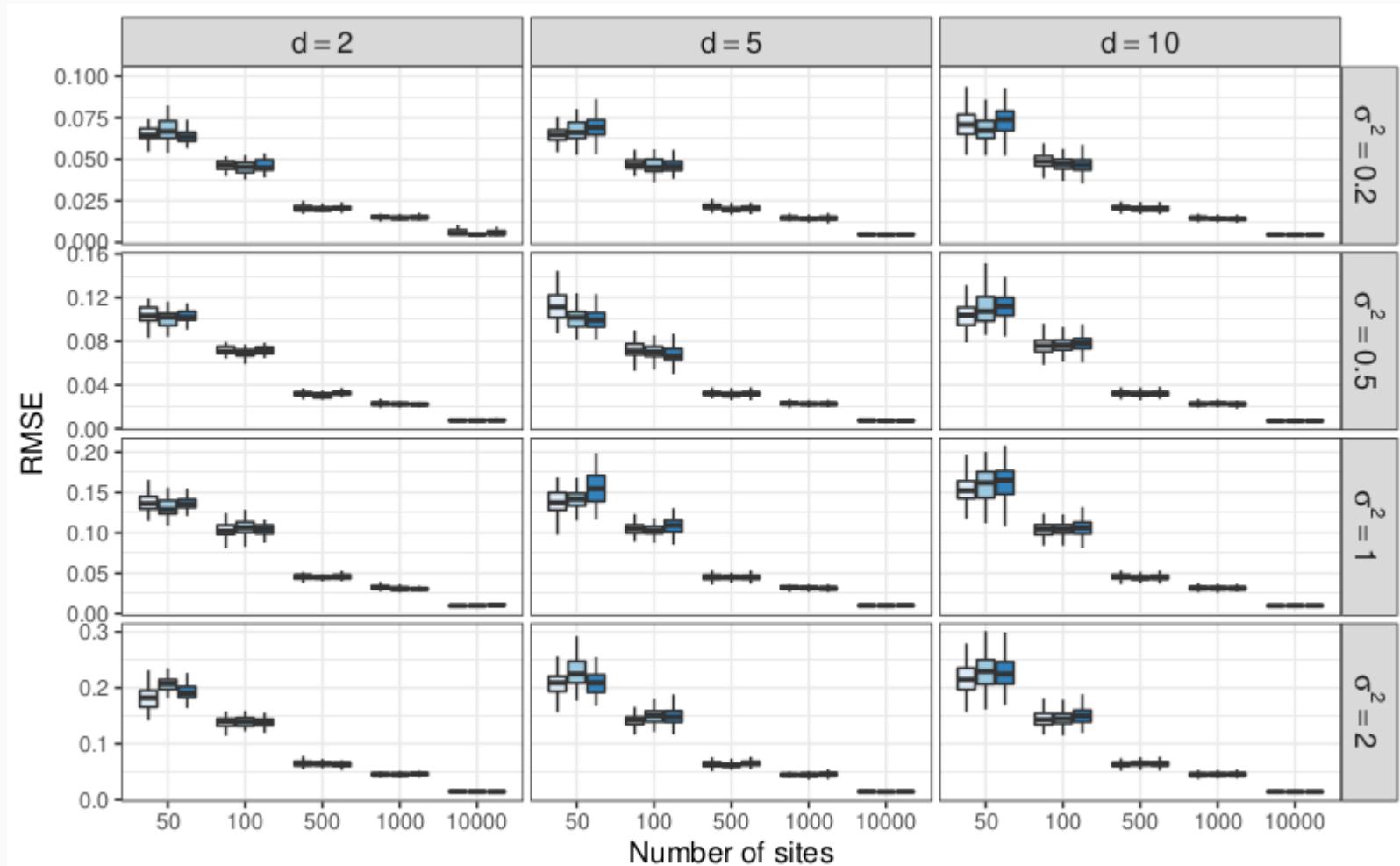
- number of samples  $n \in \{50, 100, 500, 1000, 10000\}$
- number of species/genes  $p \in \{20, 200\}$
- number of covariates  $d \in \{2, 5, 10\}$
- sampling effort  $TSS \in \{\text{low, medium, high}\}$  ( $\approx 10^4$ ,  $10^5$  and  $10^6$ )
- noise level  $\sigma^2 \in \{0.2, 0.5, 1, 2\}$
- $\Sigma$  as  $\sigma_{jk} = \sigma^2 \rho^{|j-k|}$ , with  $\rho = 0.2$
- $\Theta$  with entries sampled from  $\mathcal{N}(0, 1/d)$

# Bias of $\hat{\Theta}$



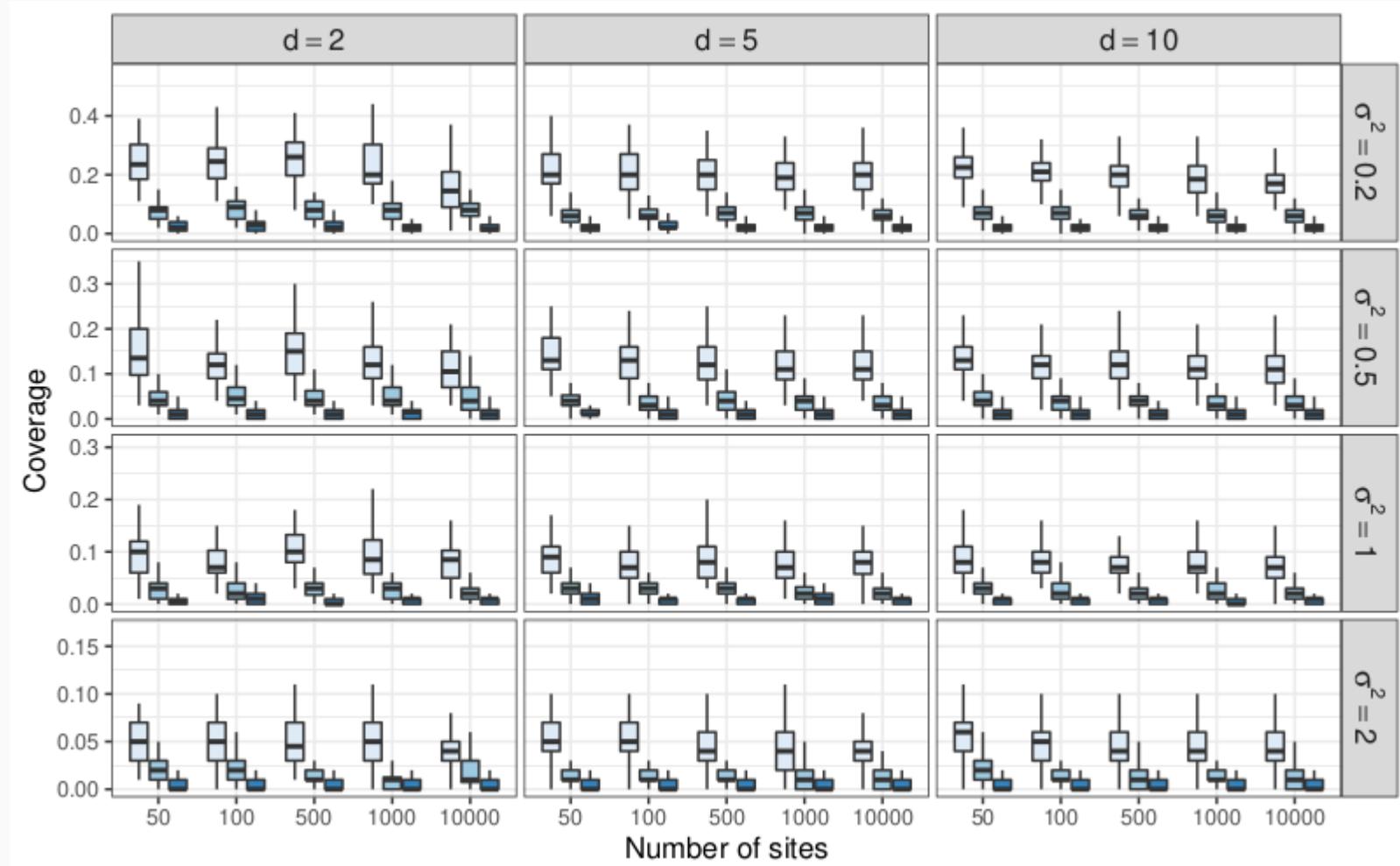
~~ asymptotically unbiased

# Root mean square error of $\hat{\Theta}$



~~ asymptotically unbiased

# 95% confident interval - Coverage



variance underestimated, no trusted confidence intervals can be derived out-of-the box

# Other ideas

## M-estimation (with M. Mariadassou)

We can derive asymptotic behavior but V-EM stationary point is not a log-likelihood stationary point, see [WM15]  $\rightsquigarrow$  Use sandwich estimator, correction.

## Exact likelihood maximization (with B. Batardière, J. Kwon)

To compare and assess at least empirically the performance of the VE-M estimator

- Use importance sampling to estimate the likelihood:

$$p_\theta(Y_i) = \int \tilde{p}_\theta(Y_i|Z)p(Z)dZ = \int \tilde{p}_\theta(Z)dZ \approx \frac{1}{n_s} \sum_{k=1}^{n_s} \frac{\tilde{p}_\theta(V_k)}{g(V_k)}, \quad (V_k)_{1 \leq k \leq n_s} \stackrel{iid}{\sim} g$$

- Estimate the gradients of the log-likelihood by plug-in:

$$\nabla_\theta \log p_\theta(Y_i) \approx \nabla_\theta \log \left( \frac{1}{n_s} \sum_{k=1}^{n_s} \frac{\tilde{p}_\theta^{(u)}(V_k)}{g(V_k)} \right)$$

# Oaks powdery mildew data set overview

Study effects of the pathogen *E.Aphiltoïdes* (mildew) wrt bacterial and microbial communities

## Species Abundances

- Microbial communities sampled on the surface of  $n = 116$  oak leaves
- Communities sequenced and cleaned resulting in  $p = 114$  OTUs (66 bacteria, 48 fungi).

## Covariates and offsets

Characterize the samples and the sampling, most important being

- tree : Tree status with respect to the pathogen (susceptible, intermediate or resistant)
- distToGround : Distance of the sampled leaf to the base of the ground
- orientation : Orientation of the branch (South-West SW or North-East NE)
- readsTOTfun : Total number of ITS1 reads for that leaf
- readsTOTbac : Total number of 16S reads for that leaf

# Abundance table (I)

```
data(oaks)
oaks$Abundance %>% as_tibble() %>%
  dplyr::select(1:10) %>%
  head() %>% DT::datatable(fillContainer = FALSE)
```

Show  entries

Search:

|   | b_OTU_1045 | b_OTU_109 | b_OTU_1093 | b_OTU_11 | b_OTU_112 | b_OTU_1191 |    |
|---|------------|-----------|------------|----------|-----------|------------|----|
| 1 | 0          | 0         | 0          | 6        | 146       |            | 1  |
| 2 | 0          | 0         | 0          | 0        | 0         |            | 1  |
| 3 | 0          | 0         | 0          | 2        | 0         |            | 0  |
| 4 | 0          | 0         | 0          | 1        | 1         |            | 1  |
| 5 | 0          | 0         | 0          | 4        | 1         |            | 1  |
| 6 | 0          | 0         | 0          | 77       | 2         |            | 20 |

Showing 1 to 6 of 6 entries

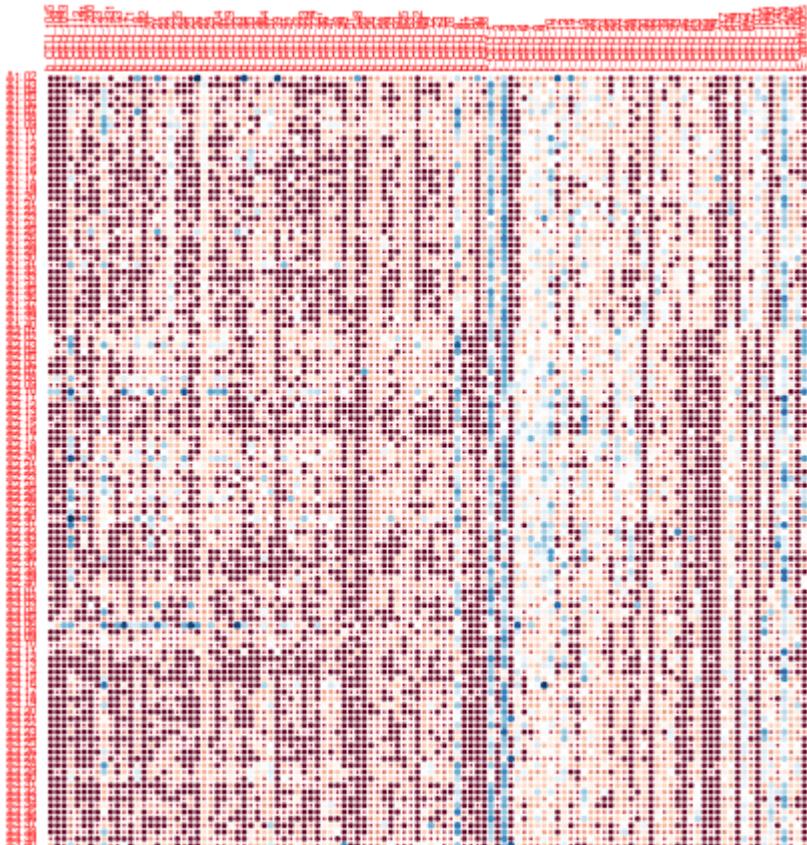
Previous

1

Next

# Abundance table (II)

```
log(1 + oaks$Abundance) %>%  
  corrplot::corrplot(is.corr = FALSE,  
    addgrid.col = NA, tl.cex = .5, cl.pos = "n")
```



# PLN with offsets and covariates (1)

## Offset: modeling sampling effort

The predefined offset uses the total sum of reads, accounting for technologies specific to fungi and bacteria:

```
M01_oaks ← PLN(Abundance ~ 1 + offset(log(Offset)) , oaks)
```

## Covariates: tree and orientation effects ('ANOVA'-like)

The `tree` status is a natural candidate for explaining a part of the variance.

- We chose to describe the tree effect in the regression coefficient (mean)
- A possibly spurious effect regarding the interactions between species (covariance).

```
M11_oaks ← PLN(Abundance ~ 0 + tree + offset(log(Offset)), oaks)
```

What about adding more covariates in the model, e.g. the orientation?

```
M21_oaks ← PLN(Abundance ~ 0 + tree + orientation + offset(log(Offset)), oaks)
```

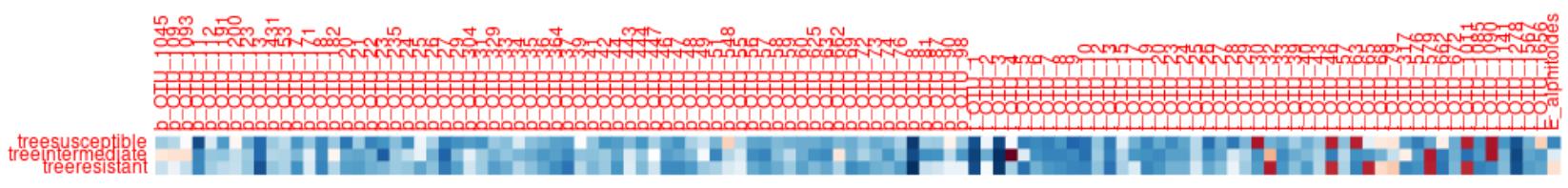
# PLN with offsets and covariates (2)

There is a clear gain in introducing the tree covariate in the model:

```
rbind(M01 = M01_oaks$criteria,
      M11 = M11_oaks$criteria, M21 = M21_oaks$criteria) %>%
knitr::kable(format = "html")
```

|     | <b>nb_param</b> | <b>loglik</b> | <b>BIC</b> | <b>ICL</b> |
|-----|-----------------|---------------|------------|------------|
| M01 | 6669            | -32252.14     | -48102.98  | -52169.64  |
| M11 | 6897            | -31524.16     | -47916.91  | -51644.03  |
| M21 | 7011            | -31438.58     | -48102.29  | -51727.13  |

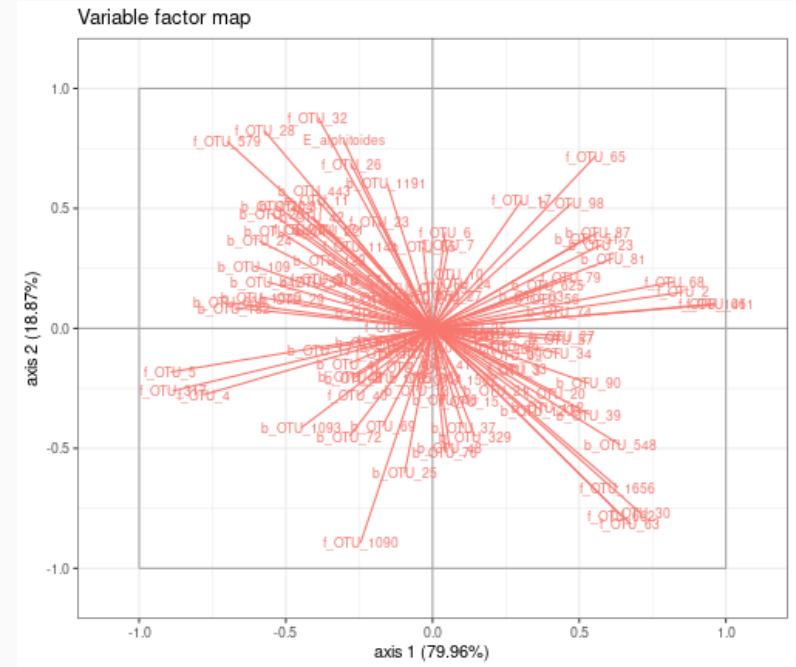
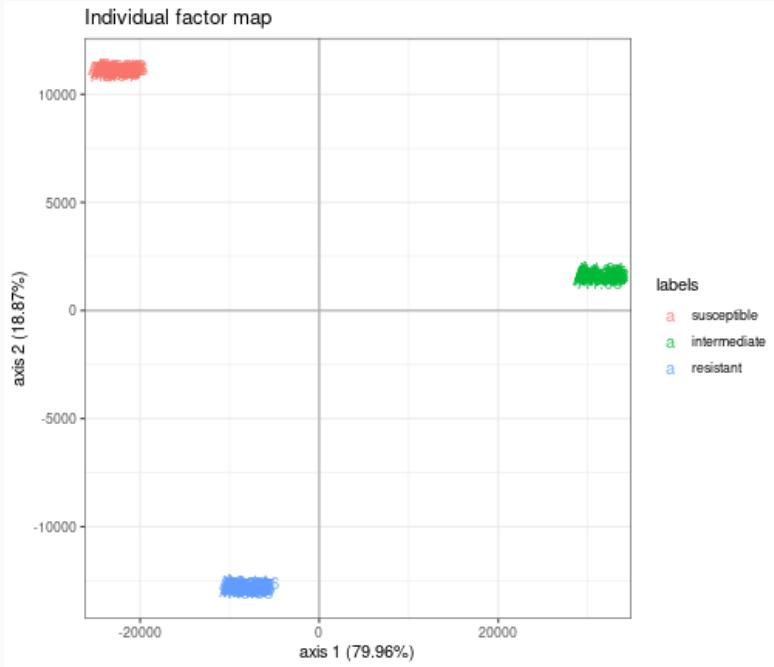
Looking at the coefficients  $\Theta$  associated with `tree` bring additional insights:



# Discriminant Analysis

Use the `tree` variable for grouping (`grouping` is a factor of group to be considered)

```
myLDA_tree <-  
  PLNLDA(Abundance ~ 1 + offset(log(Offset)), grouping = tree, data = oaks)
```

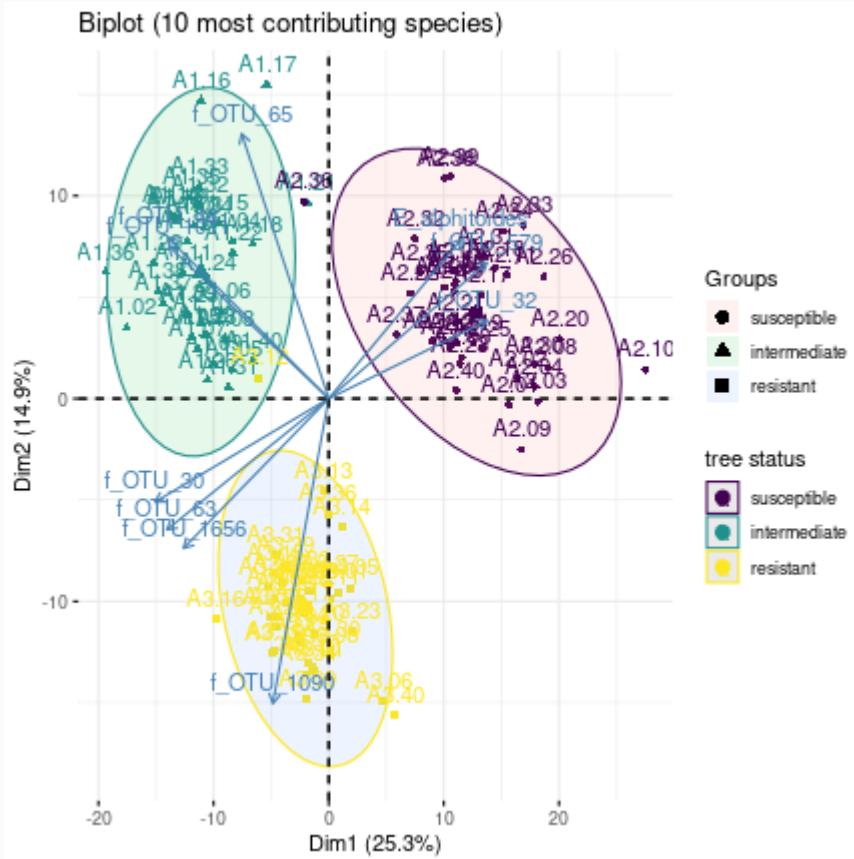


# A PCA analysis of the oaks data set

```

PCA_offset ←
  PLNPCA(Abundance ~ 1 + offset(log(Offset)), data = oaks, ranks = 1:30)
PCA_offset_BIC ← getBestModel(PCA_offset, "BIC")

```

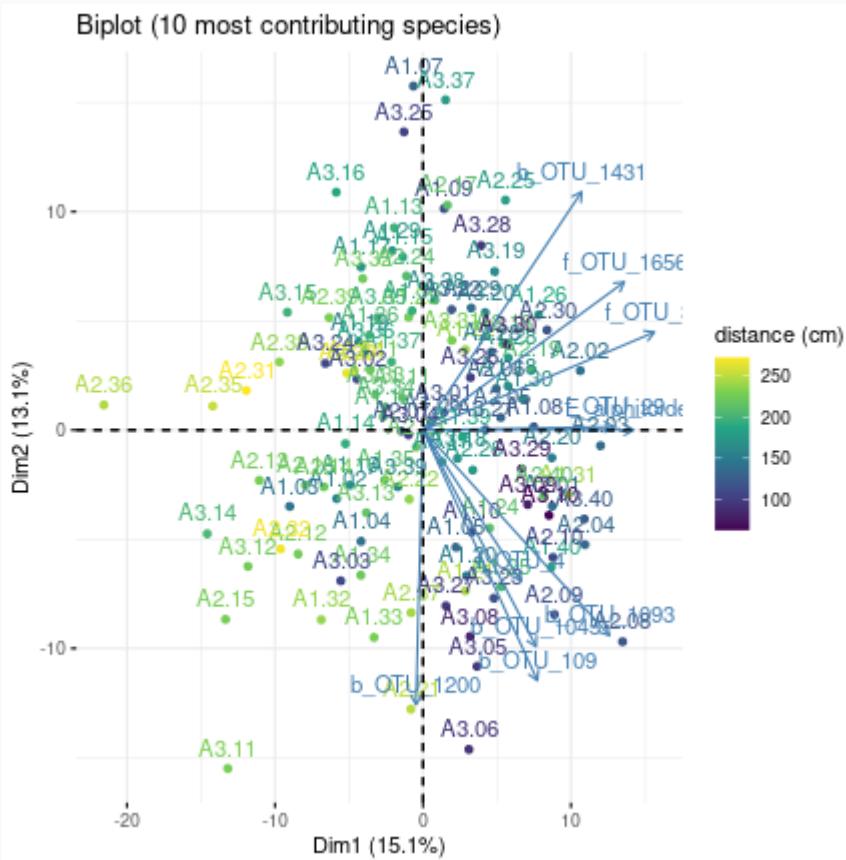


# PCA: removing covariate effects

To hopefully find some hidden effects in the data, we can try to remove confounding ones:

```
PCA_tree ←
```

```
PLNPCA(Abundance ~ 0 + tree + offset(log(Offset)), data = oaks, ranks = 1:30)
```



# Clustering of the oaks samples

```
PLN_mixtures ←  
  PLNmixture(Abundance ~ 1 + offset(log(Offset)), data = oaks, clusters = 1:3)  
myPLN_mix ← getModel(PLN_mixtures, 3)
```

```
myPLN_mix$plot_clustering_pca()
```

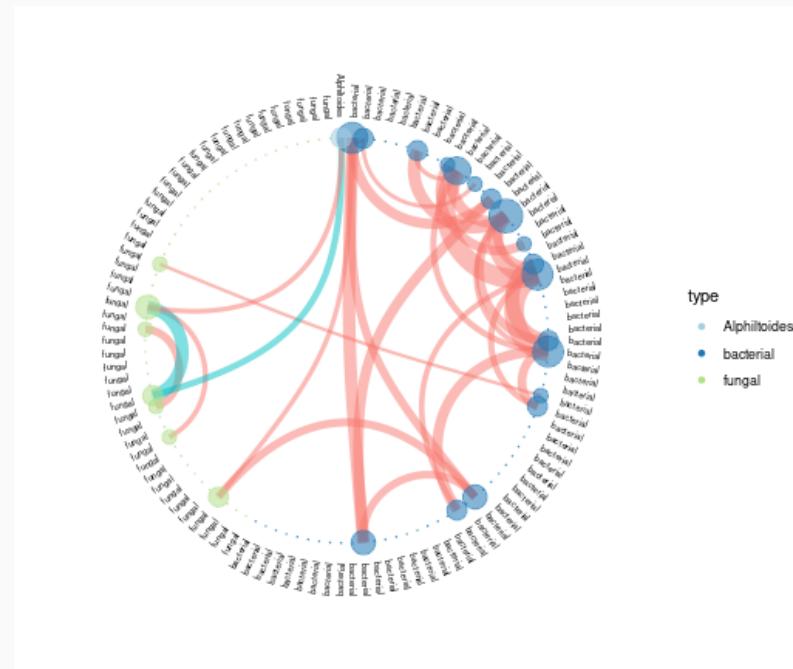
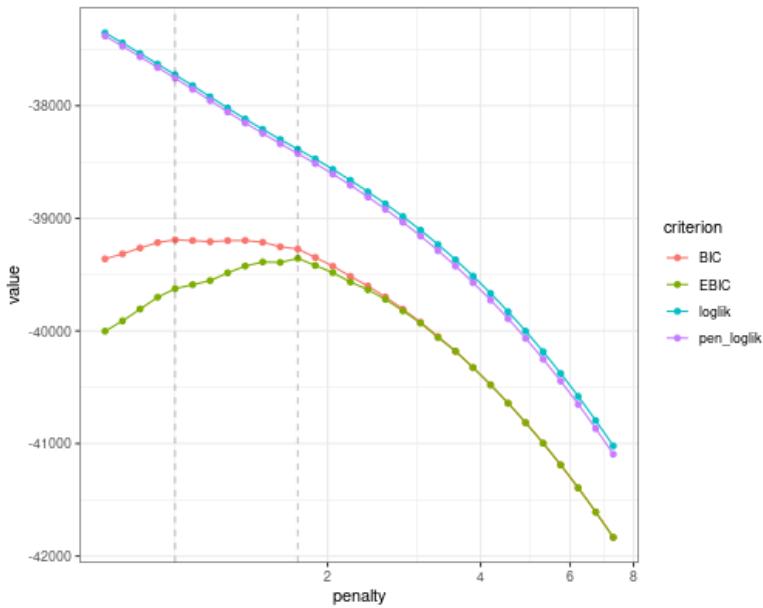
```
myPLN_mix$plot_clustering_data()
```

# Network inference

```
networks ← PLNnetwork(Abundance ~ 0 + tree + offset(log(Offset)), data = oaks)
```

Model selection criteria

Higher is better



# Conclusion

## Summary

- PLN = generic model for multivariate count data analysis
- Flexible modeling of the covariance structure, allows for covariates
- Efficient V-EM algorithm

## Extensions

- Other variants
  - zero inflation (data with *a lot* of zeros)
  - covariance structures (spatial, time series, ...)
  - Variable selection ( $\ell_1$ -penalty on the regression coefficients)
- Other models
  - Bernoulli/multinomial counterpart to PLN
  - functional data
  - multiple-data integration (e.g., Bernoulli + Poisson)

# References

- Aitchison, J. and C. Ho (1989). "The multivariate Poisson-log normal distribution". In: *Biometrika* 76.4, pp. 643-653.
- Chiquet, J., M. Mariadassou, and S. Robin (2018). "Variational inference for probabilistic Poisson PCA". In: *The Annals of Applied Statistics* 12, pp. 2674-2698. URL: <http://dx.doi.org/10.1214/18-AOAS1177>.
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- Chiquet, J., M. Mariadassou, and S. Robin (2021). "The Poisson-Lognormal Model as a Versatile Framework for the Joint Analysis of Species Abundances". In: *Frontiers in Ecology and Evolution* 9. DOI: [10.3389/fevo.2021.588292](https://doi.org/10.3389/fevo.2021.588292).
- Facon, B., A. Hafsi, M. C. de la Masselière, et al. (2021). "Joint species distributions reveal the combined effects of host plants, abiotic factors and species competition as drivers of species abundances in fruit flies". In: *Ecological Letters*. DOI: [10.1111/ele.13825](https://doi.org/10.1111/ele.13825).